TRITERPENES FROM BULBS OF *MUSCARI COMOSUM*, 3. THE STRUCTURE OF TWO MINOR NOVEL NORTRITERPENE COMPONENTS¹

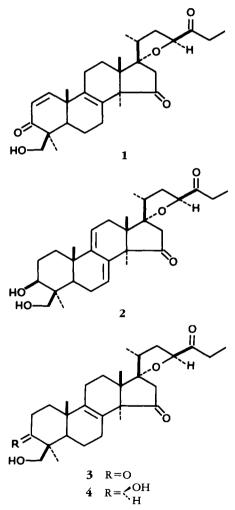
M. ADINOLFI, G. BARONE, R. LANZETTA, G. LAONIGRO, L. MANGONI, and M. PARRILLI*

Istituto di Chimica Organica e Biologica della Università di Napoli, Via Mezzocannone 16, 80134 Napoli, Italy

We recently described the structural elucidation of five components of the triterpene mixture extracted from the Muscari bulbs of comosum Mill. (Liliaceae), all characterized by a spirofused tetrahydrofuran ring in their norlanostane skeleton (1, 2). We also pointed out (1) the presence of two minor components: the first, 1, isolated from tlc fraction *a* of the mixture and the other, 2, from tlc fraction b. This paper describes the determination of the structures of these two novel components.

Compound 1, mp 181-183°, possesses the molecular formula C₂₉H₄₀O₅ (hrms). The pmr spectrum of 1 (Table 1) resembles that of compound 3, a previously (1,3) characterized component of the same triterpene fraction. The most relevant feature of the spectrum of 1 as compared to that of 3 is the presence of the signals of two vinyl protons whose chemical shift and multiplicity (δ 7.313) and δ 5.959, ABq, J = 10.56 Hz) suggest that in ring A of 1 there is a Δ^{1} -3-keto system (4,5). Accordingly, the uv spectrum displayed a maximum at 239 nm typical of steroid and terpenoid cyclohexen-3-one systems (6), and the ir spectrum showed the α,β -unsaturated ketone band at 1665 cm⁻¹. Conclusive evidence in favor of structure 1 was gained by the remarkably close similarity of the cmr spectra of 1 (Table 2) and 3(3), the signals of A-ring carbons excepted. The C=C-C=O carbon resonances were found at δ 157.13, 126.53, and 206.07, respectively (7-9). In addition, the C-4 and C-5 resonances were shifted upfield and the C-10 resonance

downfield as a consequence of the introduction of the 1,2-double bond.²



Compound 2, mp 221-224°, possesses the molecular formula $C_{29}H_{42}O_5$ (hrms). Uv absorption [EtOH, λ max 236, 241, and 250 (shoulder) nm] immediately suggests (10) the presence of the $\Delta^{7,9(11)}$ diene chromophore. The

¹Part 2: reference 2.

²Cf. Compound A (8) and azadiradion (9).

	Compound	Protons								
		18-H3	19-H ₃	30-Н ₃	32-H ₃	21-H ₃	31-H ₂	3-H		
1 2		0.972s 0.836s	1.234s 0.944s	1.321s 1.262s	1.402s 1.395s	1.145d $J_{20,21} = 6.64$ 1.211d $J_{20,21} = 6.62$	ABq $J_{AB} = 10.96$ 3.470	3.47 ^b		
_	·····	20-H	22-H,	23-Н	25-H ₂	26-H ₃	16-H ₂	Others		
			-		-	5		Others		
1		2.30m	1.950dd ^c	4.694t ^d	$\begin{array}{c} 2.487 \mathbf{q} & 1.075 \mathbf{t} \\ J_{25,26} = 7.43 \end{array}$		2.224 2.823 ABq $J_{AB} = 19.17$	7.313(1-H) 5.959(2-H) ABq $J_{1,2} = 10.56$		
2		2.3m	1.95m	4.684t ^d	2.501q J _{25,2}	1.078t 26=7.35	2.175 2.798 ABq $J_{AB} = 19.12$	5.325m (11-H) 6.721bd (7-H)		

TABLE 1. Pmr (270 MHz) Chemical Shifts in CDCl₃^a

^aAll chemical shift values are given in δ (ppm) relative to TMS. Coupling constants are given in Hz and were inferred from pertinent decoupling experiments. Assignment of the methyl resonances was based mainly on comparison with the spectra of **3** and **4** (2).

^bBuried within the 31-H₂ signal.

^cAA' part of an AA'X system, further split (J=3.13) by coupling with 20-H. ^dX part of an AA'X system ($J_{AX}+J_{A'X}=18.00$).

TABLE 2. Chirt (67.86 MHz) Chemical Shifts in CDCI ₃ of T											
C-1	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10		
157.13	126.53	206.07	48.72	47.35	18.03	25.52	135.02	135.02	40.43		
C-11	C-12	C-13	C-14	C-15	C-16	C-17	C-18	C-19	C-20		
21.41	23.26	47.65	57.73	214.37	51.53	90.86	20.32	19.20	43.30		
C-21	C-22	C-23	C-24	C-25	C-26	C-30	C-31	C-32			
17.08	36.66	81.55	211.70	32.29	7.29	21.88	64.90	24.23			

TABLE 2. Cmr (67.88 MHz) Chemical Shifts in CDCl₃ of 1^a

^aChemical shift values are given in δ (ppm) relative to TMS. The assignments of the signals are based on the comparison with the spectrum of **3** (3).

pmr spectrum (Table 1), resembling that of 4 (2), supports this structural feature: 11-H and 7-H resonate at δ 5.325 and δ 6.721, respectively. The displacement to lower field of the signal of 7-H with respect to other lanosta-7,9(11)- dienes (11) can be accounted for by the effect of the 15-keto group, since a similar chemical shift value has been reported (12) for the 7-H of another 15keto lanosta-7,9(11)-diene. In spite of the absence of supporting cmr evidence, due to the lack of a sufficient amount of material, structure 2 was readily confirmed by converting 4 into 2 by reaction with m-chloroperbenzoic acid followed by treatment with 80% aqueous HOAc, according to a simple modification of the well known procedure for the transformation of Δ^8 steroids and terpenoids into the $\Delta^{7,9(11)}$ analogues (13).

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.— See reference 1.

ISOLATION PROCEDURE.—Nortriterpenes 1 and 2 were obtained from fresh fractions a and b, respectively, isolated by the procedure described in reference 1 from the bulbs of *M. comosum*.

Compound 1 was the minor component isolated by tlc from the mother liquors of the crystallization of fraction a (1). It had mp 181-183° (from Me₂CO); ms m/z 468.2863 (M⁺, calcd. for $C_{29}H_{40}O_5$ 468.2876), 438 (base peak), 411 (M⁺-57, CH₃CH₂CO), 380, 268, 241; ir (CHCl₃): 3300 (OH), 1735 (five ring C=O), 1715 (C=O), 1685 (C=C-C=O) cm⁻¹; uv (EtOH), λ max 239 nm; pmr Table 1; cmr Table 2.

Compound 2 was the minor component isolated by tlc from the fraction b(1). It had mp 221-224° (from MeOH); ms m/z 470.3023 (M⁺, calcd. for C₂₉H₄₂O₅ 470.3032), 413, 324 (base peak); ir (CHCl₃): 3300 (OH), 1735 (five ring C=O), 1715 (C=O) cm⁻¹; uv (EtOH), λ max 236, 241, 250 (shoulder) nm; pmr Table 1.

CONVERSION OF 4 INTO 2.—A solution of 4 (1) (3 mg) and *m*-chloroperbenzoic acid (4 mg) in CHCl₃ (2 ml) was allowed to stand at room temperature for 40 h. The solution was then treated with solid KI (traces), acidified by addition of HOAc, washed with 0.1 N sodium metabisulphite, with 2 N NaOH and finally with H₂O, and evaporated to dryness. The residue was dissolved in 80% aqueous HOAc (2 ml) and allowed to stand at room temperature for two days. The reaction mixture was diluted with CHCl₃, washed with saturated aqueous NaCl and evaporated. Tlc (silica gel; 8:2, $EtOAc-CHCl_3$) of the residue gave a solid (2 mg) which was identical (tlc, pmr) to the sample of **2** isolated from the bulbs as above.

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